

# Deficits in Bone Geometry in Growth Hormone-Deficient Prepubertal Boys Revealed by High-Resolution Peripheral Quantitative Computed Tomography

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## Keywords

Growth hormone deficiency · Bone · HR-pQCT · Pediatric endocrinology

## Abstract

**Introduction:** Although growth hormone (GH) is essential for attainment of peak bone mass, bone health in prepubertal children with GH deficiency is not routinely evaluated. The objective of this study was to evaluate bone microarchitecture in GH-deficient (GHD) boys using high-resolution peripheral quantitative computed tomography (HR-pQCT). **Methods:** Fifteen control and fifteen GHD, GH naïve prepubertal boys were recruited for a case-control study at a major academic center. Subjects with panhypopituitarism, chromosomal pathology, chronic steroids, or stimulant use were excluded. Volumetric bone mineral density (vBMD; total, cortical, and trabecular), bone geometry (total, cortical and trabecular cross-sectional area, cortical perimeter), bone microarchitecture, and estimated bone strength of the distal radius and tibia were assessed by HR-pQCT. Areal BMD and body composition were assessed by DXA. Insulin-like growth factor 1 (IGF-1), osteocalcin, C telopeptide, and P1NP levels

were measured. **Results:** GHD subjects had a significantly smaller cortical perimeter of the distal radius compared to controls ( $p < 0.001$ ), with the difference in cortical perimeter persisting after adjusting for height z score, age, lean mass, and 25-hydroxyvitamin D level ( $p < 0.05$ ). No significant differences were found in vBMD. No significant differences were found in microarchitecture, estimated strength, areal BMD, body composition, or bone turnover markers. Analysis showed significant positive correlations between IGF-1 levels and cortical parameters. **Discussion/Conclusions:** Prepubertal GHD boys had deficits in bone geometry not evident with DXA. Larger prospective/longitudinal HR-pQCT studies are needed to determine the extent of these deficits, the need for routine bone evaluation, and the timing of GH replacement for prevention or restoration of these deficits.

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## Introduction

Although growth hormone (GH) is responsible for longitudinal bone growth, it also plays an important role in building and maintaining bone mineral density (BMD)

and in altering bone architecture throughout life [1, 2]. GH, by acting directly and by stimulating insulin-like growth factor 1 (IGF-1), is essential for achieving peak bone mass, and contributes to mitigating the risk of osteoporosis and subsequent fracture in the future [2–5]. GH deficiency during childhood and puberty may compromise accrual of bone mass and formation of normal bone architecture because a significant amount of bone mass is achieved by the end of puberty, with peak bone mass achieved during the late second to early third decade of life [3, 6]. Even though GH-deficient (GHD) prepubertal children may not be at risk for fractures, studies have shown that untreated children with GH deficiency (mean age 7–11.7 years) have lower dual X ray absorptiometry (DXA) measures of bone mineral apparent density (BMAD, as an estimate of volumetric BMD) of the lumbar spine, radius, and total body [2, 7–9], while untreated adults with GH deficiency have increased fracture risk as well as lower BMD scores [10–12].

Prior studies in GHD children have evaluated BMD using DXA, though this method has significant disadvantages. Measurements are affected by size, thereby underestimating BMD in people with smaller stature [13]. Additionally, DXA only measures areal BMD from a 2-D projection of bone and does not measure actual vBMD (volumetric BMD). Resolution of the images is low, and DXA cannot be used to assess BMD in the cortical and trabecular bone separately or assess microarchitectural characteristics that also contribute to bone strength [14–16].

High-resolution peripheral quantitative computed tomography (HR-pQCT) is a state-of-the-art imaging technology that can scan human subjects in vivo at resolutions high enough to enable characterization and evaluation of volumetric density, geometry, and microarchitecture. As the resolution is high (voxel size 61  $\mu\text{m}$ ), it even allows for segmentation of trabecular and cortical bone and their respective evaluation. Bone microarchitecture and bone density contribute to increased bone strength [17, 18] which can decrease risk of fracture in children [19, 20]. Knowledge regarding the effects of GH deficiency on skeletal health is limited though because bone parameters are not routinely evaluated in young children with GH deficiency. A recent HR-pQCT study in adults with childhood-onset GH deficiency who were no longer receiving GH replacement showed deficits in vBMD, architecture, and bone strength [21]. Schweizer et al. [22] performed the only study evaluating trabecular and cortical bone compartments in the pediatric GHD population, using a lower-resolution imaging technique (peripheral QCT) with the inability to evaluate microar-

chitectural bone features. Additionally, studies using HR-pQCT in healthy children tend to focus on the adolescent or peripubertal populations [23, 24]. Proper characterization of bone health in young, prepubertal GHD children has significant implications for a critical window of intervention which can impact long-term bone health.

The primary goal of this study was to use HR-pQCT to evaluate bone geometry, vBMD, bone architecture, and bone strength in GHD pre-pubertal boys compared to healthy control subjects. The relationship between GH levels, IGF-1 levels, bone markers, and HR-pQCT measures was also assessed in the GHD cohort.

## Materials and Methods

### *Participants*

A total of 30 boys, 5–11 years of age were recruited between 2016 and 2018 from the general pediatric practices and the pediatric endocrinology practice associated with Columbia University Medical Center, as well as other nearby practices. Informed consent was obtained from a parent or legal guardian, and assent was obtained for children >7 years of age. The study was approved by the Institutional Review Board of Columbia University Medical Center. All participants were assessed as prepubertal based on evaluation of testicular size <4  $\text{cm}^3$  and had no disabilities that would limit normal physical activity. Fifteen boys had isolated GHD and were naive to therapy. Along with height and growth velocity measurements, the diagnosis of GHD was reconfirmed with a peak serum GH level <10 ng/mL in response to 2 stimulation tests (clonidine/arginine, glucagon, or arginine/L-dopa) [25]. All bone age (BA) studies were documented and assessed by the pediatric endocrinologist and the radiologist. Standards of Greulich and Pyle were used to estimate BA. Fifteen subjects with heights between the 3rd and 97th percentiles who were healthy, with no concerns regarding height or growth velocity served as controls and were recruited from the general pediatric practices affiliated with Columbia as well as other nearby practices. Control and GHD subjects were not matched for height, as that would have required including much younger control subjects. Neither control nor GHD participants had chronic health issues (beyond GHD) that would interfere with bone health, and all were ambulatory. Children with panhypopituitarism, chromosomal diagnosis, or on chronic medication, including levothyroxine, systemic or inhaled steroids, or stimulants, were not included. Use of intermittent antihistamines and vitamin supplementation was allowed. All children were born at appropriate size for gestational age. Nutritional history was not formally evaluated. Ethnicity was determined based on parental self-report. In the control group, there were 9 Hispanic children, 4 Caucasian children, 1 African-American child, and 1 Asian child. In the GHD group, there were 7 Hispanic children, 4 Caucasian children, 1 African child, 2 Asian children, and 1 Middle Eastern child.

### *Anthropometric Data Acquisition*

Standing heights and weights were measured using a wall-mounted stadiometer and electronic scale, with participants dressed in light clothing. To determine growth velocity from med-

**Table 1.** Subjects' characteristics

|  | Control ( <i>n</i> = 15) | GHD ( <i>n</i> = 15) | <i>p</i> value |
|--|--------------------------|----------------------|----------------|
| <i>Demographics</i>                          |                          |                      |                |
| Height <i>Z</i> score                        | -0.39±1.0                | -2.27±0.61           | <0.001*        |
| Tibial length, mm                            | 271.67±27.88             | 249.33±21.20         | <0.001*        |
| Radius length, mm                            | 183.28±22.09             | 173.8±16.48          | 0.469          |
| Age, years                                   | 7.82±1.32                | 8.85±1.21            | 0.035*         |
| Hispanic ethnicity (%)                       | 9 (60)                   | 7 (47)               | 0.714          |
| BMI %ile                                     | 57.27±27.3               | 38.61±28.21          | 0.097          |
| 25-hydroxyvitamin D, ng/mL                   | 25.62±6.39               | 22.4±8.36            | 0.251          |
| IGF-1, ng/mL                                 | n/a                      | 140.93±66.57         | n/a            |
| Peak growth hormone level, ng/mL             | n/a                      | 5.62±2.25            | n/a            |
| <i>Bone turnover markers</i>                 |                          |                      |                |
| CTX, ng/mL                                   | 1.63±0.42                | 1.72±0.52            | 0.702          |
| OC, ng/mL                                    | 116.61±51.59             | 98.81±46.36          | 0.338          |
| PINP, µg/mL                                  | 572.87±146.39            | 489.99±162.36        | 0.160          |
| <i>DXA</i>                                   |                          |                      |                |
| Whole body (subtotal) BMD, g/cm <sup>2</sup> | 0.61±0.06                | 0.57±0.06            | 0.088          |
| Whole body BMD <i>Z</i> score                | 0.14±0.75                | -0.29±0.65           | 0.109          |
| AP spine BMD, g/cm <sup>2</sup>              | 0.54±0.07                | 0.51±0.06            | 0.255          |
| AP spine BMD <i>Z</i> score                  | 0.15±1.06                | 0.15±0.84            | 0.999          |
| 1/3 forearm BMD, g/cm <sup>2</sup>           | 0.48±0.05                | 0.46±0.43            | 0.229          |
| 1/3 forearm BMD <i>Z</i> score               | 0.36±1.37                | 0.11±0.81            | 0.547          |
| R arm area, cm <sup>2</sup>                  | 102.07±12.59             | 95.69±11.70          | 0.180          |
| Fat mass, g                                  | 6,718.73±2,738.89        | 5,919.33±2,133.86    | 0.290          |
| Lean mass, g                                 | 17,827.72±3,306.08       | 15,778.33±2,992.67   | 0.086          |
| Percent fat                                  | 25.71±5.73               | 26.17±6.38           | 0.839          |
| Visceral adipose tissue, g                   | 151.33±67.13             | 146.73±44.29         | 0.827          |

Summary statistics for demographics, bone turnover markers, and DXA. Values shown as mean ± SD or *n* (%); *p* values generated by two-sample *t* test or  $\chi^2$  test; \* *p* value <0.05.

ical records in GHD subjects, height measurements were taken at intervals of 4–6 months. Radial length was assessed as the distance from the olecranon to the ulnar styloid process, measured medially with elbow flexed at a 90-degree angle and palm facing inward. Tibial length was assessed as the distance from the medial malleolus to medial tibial plateau, measured with the knee flexed at a 90-degree angle to the floor [24].

#### Calculated vBMD and Body Composition Acquisition Using DXA

DXA scans were obtained at the Body Composition Unit of Columbia University Medical Center. DXA scans of the whole body excluding head, posteroanterior lumbar spine (L1–L4) and right forearm were obtained using Hologic QDR 4500 in array mode (Hologic Inc., Waltham, MA, USA). Scans were analyzed using APEX 4.5.3 software and read by 2 certified densitometrists in the division of pediatric endocrinology (A.S. and I.F.) for quality assurance. Height correction was performed for all subjects [26].

#### Bone Architecture, vBMD and Strength Acquisition with HR-pQCT

Trabecular and cortical vBMD and microarchitecture were assessed using HR-pQCT (XtremeCT-II, Scanco Medical, Brüt-

tisellen, Switzerland). Each subject's right radius and tibia were scanned and placed in a carbon-fiber cast to minimize limb motion during scan acquisition. If the subject had a history of fracture in the right limb, then the left limb was scanned. A standard anteroposterior scout view was taken (fixed settings of the machine) to assess the growth plate and then place the reference line at the most proximal end of the growth plate in order to ensure that the growth plate was not irradiated, although the radiation dosage was very minimal (<5 µSv/scan). A 10.2-mm scan region comprising of 168 slices with an isotropic voxel size of 61 µm was acquired at both sites. The scans were acquired at a relative offset to the most proximal slice from the reference line; the offset being 4.5 and 7% of the limb length at radius and tibia, respectively. Due to possible differences in height and limb length in these growing children, it was crucial to use a relative offset in order to scan the same or similar regions of interest across the cohort. This relative offset ensured that the same scan region was obtained despite varying limb lengths. Scans were performed using the standard manufacturer in vivo imaging protocol [27, 28]. All scans were assessed for motion on a scale of 1–5 with 1 indicating no motion and 5 indicating significant motion [29], and scans with motion scores >3 were excluded from the analysis (1 radius, 1 tibia). Scans were obtained and analyzed by a single qualified technician in order to minimize interoperator variability.

Finite element analysis (FEA) was performed using the HR-pQCT images to estimate whole bone stiffness (N/mm) and failure load (N) [30, 31]. Uniaxial compression was simulated to 1% strain using a homogeneous Young's modulus of 6,829 MPa and Poisson's ratio of 0.3 [32] to estimate stiffness. Failure load (FL) was estimated based on the criterion by Pistoia et al. [33]. We used a commercial FE solver (FAIM, v7.1; Numerics88, Calgary, AB, Canada) on a desktop workstation (Linux CentOS 7.1, 2 × 6-core Intel Xenon, 64 GB RAM) to solve the models.

The CV (%) (coefficient of variation) for DXA sites at our center is LS (lumbar spine) <1% FN (femoral neck) <1.5%, forearm <1%, body composition 1%. For HR-pQCT (XtremeCT II) at our center, all density measures are <1% at both distal radius and tibia, microarchitecture <4.5% at both sites with the exception of Ct.Po being <16% at radius and <10% at tibia, FEA measures (stiffness and FL) <7% at radius and <3% at tibia.

#### Biochemical Assays

Fasting blood samples were obtained and stored for batch analysis. Serum osteocalcin and C telopeptide were measured by ELISA, and PINP measured by radioimmunoassay, all from Immundiagnostic Systems (Gaithersburg MD). Growth hormone and IGF-1 assays were run by IDS-iSYS Specialty Immunoassay System at the Pathology's Clinical Pharmacology and Toxicology Laboratory at the Irving Institute for Clinical and Translational Research, Columbia University Medical Center (New York, NY, USA). 25-hydroxyvitamin D<sub>2</sub> and 25-hydroxyvitamin D<sub>3</sub> were measured using ultra-performance liquid chromatography-tandem mass spectrometry by the Biomarkers Core Lab, Irving research Institute for Clinical and Translational Research, Columbia University Medical Center (New York, NY, USA).

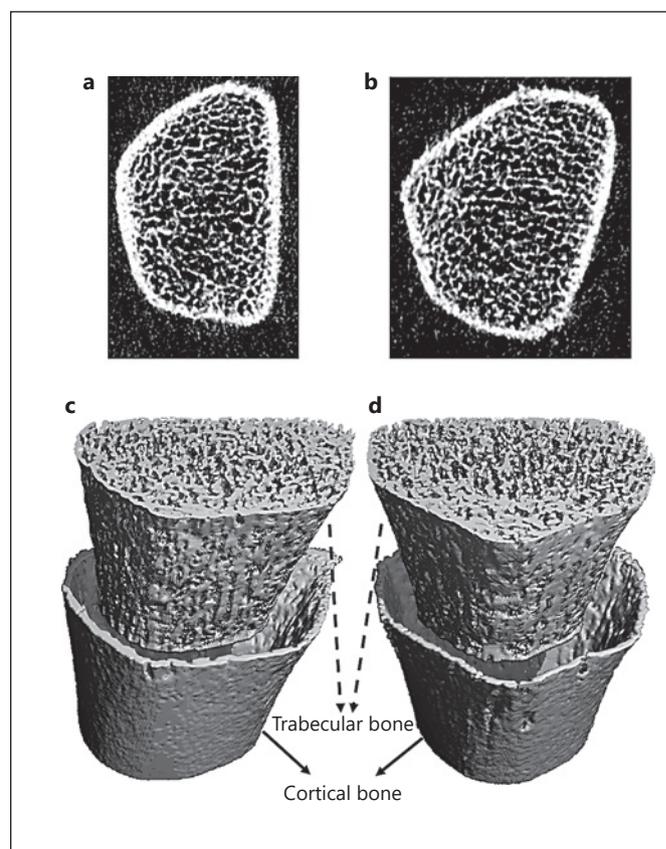
#### Statistical Methods

Descriptive statistics were used to summarize participants' characteristics by group: GHD and control subjects. Continuous measurements were expressed as mean ± standard deviation or median (interquartile range), and differences between the two groups were tested using two-sampled *t* tests or Wilcoxon rank-sum tests for measurements non-normally distributed. Linear regression models were employed to assess differences in HR-pQCT measurements between GHD and control subjects, adjusting for age, height standard deviation score, lean mass, and 25-hydroxyvitamin D level. Spearman correlation coefficients were used to quantify the strength of association between HR-pQCT parameters and bone turnover markers, GH, and IGF-1 levels, respectively. All statistical tests were two tailed, and *p* values <0.05 were considered statistically significant.

## Results

### Clinical, Anthropometric, and Biochemical Characteristics

Clinical, anthropometric and biochemical parameters characterizing our cohort are presented in Table 1. GHD prepubertal males were on average 1 year older, and as expected significantly shorter than controls (height *z* score in GHD group  $-2.27 \pm 0.61$  vs. controls  $-0.39 \pm 1$ ; *p* <



**Fig. 1.** HRpQCT scan of distal radius in growth hormone-deficient (left) and control (right) subjects. **a, b** A single cross-sectional slice of the scan showing the geometry and microstructure of the bone. **c, d** The segmented trabecular and cortical bones.

0.001). Although GHD subjects were shorter, there was no significant difference in limb length of the radius between the two groups (*p* = 0.47). Height differences were due to differences in tibial length (*p* < 0.001) and likely differences in trunk length (though not measured in this study). No significant difference in the BMI percentile was found among the groups, with actual BMI ranging between 14.0 and 19.6 kg/m<sup>2</sup> across all participants. GHD prepubertal males had IGF-1 *z* score of  $-0.23 \pm 1.26$  and a BA of  $7.2 \pm 1.8$  ( $-2.1$  standard deviations below the mean for chronological age) with an annual growth velocity of  $4.2 \pm 0.55$  cm/year (<10th percentile for age). All subjects were vitamin D sufficient. There were no significant differences between GHD and control groups in absolute values of bone formation and resorption markers.

#### DXA

No significant differences between the GHD and control groups were found in bone-projected area at the ra-

**Table 2.** Summary statistics for HR-PQCT radius.

|  | Control ( <i>n</i> = 15) | GHD ( <i>n</i> = 15) | <i>p</i> value |
|--|--------------------------|----------------------|----------------|
| <i>HR-pQCT radius</i>  |                          |                      |                |
| Total cross-sectional area, mm <sup>2</sup>  | 139.18±23.78             | 109.57±17.89         | <0.001*        |
| Cortical perimeter, mm   | 46.76±3.71               | 41.18±3.59           | <0.001*        |
| Cortical cross-sectional area, mm <sup>2</sup>   | 30.24±5.84               | 27.04±4.99           | 0.123          |
| Trabecular cross-sectional area, mm <sup>2</sup>   | 111.41±24.15             | 84.74±15.05          | <0.001*        |
| Total vBMD, mg HA/cm <sup>3</sup>  | 289.67±50.73             | 297.17±28.5          | 0.632          |
| Trabecular vBMD, mg HA/cm <sup>3</sup>   | 179.3±42.59              | 164.02±26.51         | 0.513          |
| Cortical vBMD, mg HA/cm <sup>3</sup>   | 695.74±65.41             | 731.25±26.96         | 0.076          |
| Trabecular number, 1/mm  | 1.79±0.26                | 1.66±0.16            | 0.110          |
| Trabecular thickness, mm   | 0.21±0.02                | 0.21±0.01            | 0.271          |
| Cortical thickness, mm   | 0.74±0.16                | 0.75±0.11            | 0.721          |
| Cortical porosity  | 0.01±0.001               | 0.01±0.001           | 0.947          |
| Stiffness, N/mm  | 16,578.79±5,489.55       | 14,835.87±3,677.58   | 0.321          |
| Failure load, N  | 881.79±283.96            | 778.07±200.75        | 0.263          |
| <i>HR-pQCT tibia</i>   |                          |                      |                |
| Total cross sectional area, mm <sup>2</sup>  | 524.18±66.20             | 478.99±81.71         | 0.124          |
| Cortical perimeter, mm   | 89.29±6.32               | 85.33±7.90           | 0.159          |
| Cortical cross-sectional area, mm <sup>2</sup>   | 60.48±14.41              | 51.59±10.16          | 0.088          |
| Trabecular cross-sectional area, mm <sup>2</sup>   | 468.30±65.22             | 431.79±74.92         | 0.184          |
| Total vBMD, mg HA/cm <sup>3</sup>  | 259.22±36.01             | 252.83±29.46         | 0.609          |
| Trabecular vBMD, mg HA/cm <sup>3</sup>   | 206.06±31.48             | 203.53±29.65         | 0.475          |
| Cortical vBMD, mg HA/cm <sup>3</sup>   | 684.12±33.58             | 682.07±31.07         | 0.868          |
| Trabecular number, 1/mm  | 1.84±0.19                | 1.81±0.17            | 0.696          |
| Trabecular thickness, mm   | 0.24±0.02                | 0.24±0.02            | 0.984          |
| Cortical thickness, mm   | 0.76±0.21                | 0.67±0.14            | 0.128          |
| Cortical porosity  | 0.01±0.01                | 0.01±0.01            | 0.580          |
| Stiffness, N/mm  | 59,011.38±20,135.76      | 54,980.93±21,664.78  | 0.616          |
| Failure load, N  | 3,032.08±935.43          | 2,625.93±858.27      | 0.242          |
| Values shown as mean ± SD; <i>p</i> values generated by two-sample <i>t</i> test test; * <i>p</i> value <0.05. |                          |                      |                |

dius (“R arm area”), whole body minus head, 1/3 forearm or AP spine volumetric BMD or BMD *z* score. Fat mass, lean mass, percent fat, and visceral adipose tissue did not differ significantly between the two groups (Table 1). Interestingly in GHD subjects, IGF-1 levels had a positive correlation with whole-body areal BMD ( $r = 0.73$ ,  $p = 0.003$ ) as well as lean mass ( $r = 0.76$ ,  $p = 0.002$ ). As expected in GHD subjects, peak stimulated growth hormone levels had a negative correlation with percent fat ( $r = -0.64$ ,  $p = 0.01$ ) and visceral adipose tissue ( $r = -0.66$ ,  $p = 0.01$ ).

#### *HR-pQCT Measurements at the Distal Radius and Tibia*

##### *Bone Geometry and Structure*

At the radius, GHD subjects had significantly smaller cortical perimeter indicating narrower bones, when compared to controls (Fig. 1). Trabecular cross-sectional area was on average 24% smaller, and cortical perimeter was on

average 12% smaller in GHD subjects compared to controls ( $p < 0.001$ ) (Table 2). The difference in cortical perimeter persisted after controlling for height *z* score, age, lean mass, and 25-hydroxyvitamin D level. Regression analysis showed that on average, GHD prepubertal males had 4.20 mm smaller cortical perimeter ( $p < 0.05$ ) compared to controls (Table 3). In the GHD group, IGF-1 correlated significantly with cortical parameters (cortical vBMD, cortical cross-sectional area, and cortical thickness) at the radius with coefficients ranging from  $r = 0.60$  to  $0.80$  (all  $p < 0.05$ ). None of the geometric parameters were significantly different between the groups at the tibia.

##### *Volumetric BMD, Microarchitecture, and Estimated Bone Strength*

No significant differences between the two groups were found in total, cortical, or trabecular vBMD or any of the microstructure parameters in both the radius and

**Table 3.** Regression analysis results for two different outcomes, controlling for height Z score, age, lean mass, and 25-hydroxyvitamin D level

|   | Estimate | Std. error | p value |
|---|----------|------------|---------|
| <i>Outcome: cortical perimeter</i>              |          |            |         |
| Intercept                                       | 38.79    | 4.99       | <0.0001 |
| GHD (vs. control)                               | -4.21    | 1.96       | 0.043   |
| Height Z score                                  | 1.85     | 1.06       | 0.094   |
| Age   | 1.70     | 0.99       | 0.100   |
| Lean mass                                       | -0.0001  | 0.001      | 0.724   |
| Vitamin D                                       | -0.07    | 0.08       | 0.378   |
| <i>Outcome: trabecular cross-sectional area</i> |          |            |         |
| Intercept                                       | 47.21    | 27.72      | 0.103   |
| GHD (vs. control)                               | -17.99   | 10.87      | 0.112   |
| Height Z score                                  | 7.98     | 5.87       | 0.188   |
| Age   | 8.16     | 5.50       | 0.152   |
| Lean mass                                       | -0.0001  | 0.002      | 0.953   |
| Vitamin D                                       | 0.192    | 0.449      | 0.673   |

tibia. Stiffness and failure load as measures of estimated bone strength did not differ significantly between the two groups (Table 2).

## Discussion/Conclusions

This study evaluated prepubertal GHD young boys using HR-pQCT. The results of our study provide novel insights into the structural characteristics of cortical and trabecular bone architecture in prepubertal boys diagnosed with GHD using a state-of-the-art technology. Standard bone morphology in the skeleton and as observed at the radius and tibia from HR-pQCT scans is comprised of trabecular bone surrounded by cortical bone with a surrounding periosteum. Our study demonstrates that GHD prepubertal boys had bones that were narrower compared to controls after adjusting for age, height z score, lean mass, and 25-hydroxyvitamin D level. This was determined by a significantly smaller cortical perimeter, indicative of a smaller periosteal boundary. Studies have shown that differences in bone size between boys and girls likely contribute to differences in fracture risk later in life [34, 35], indicating the important role that bone size plays. Animal models with GHD have also demonstrated that GHD results in deterioration of bone size, microarchitecture, and mechanical properties [36].

In order to eliminate the effect of variation of limb length and ensure comparable regions when scanning, a relative offset from the reference line was used. This is

important as bone geometry and microarchitecture vary along the length of long bones, with the distal (epiphysis and metaphysis) region having a dense mesh of trabecular network surrounded by a thin cortical shell. Moving proximally (towards the diaphysis), the trabecular mesh dissipates giving rise to the marrow cavity surrounded by a thick cortical shell. Hence, there is a gradient in bone properties depending on the scan region. Had a fixed offset been used, the differences in scan region would have potentially caused differences in density, geometry, and microstructure measurements across subjects, thereby confounding the actual variation between the GHD and control groups. Additionally, although limb length naturally varied between subjects, a difference in length does not imply a difference in cross-sectional area. Bones can be longer or shorter (axial length of the bone) and narrower or wider (cross section of the bone) with or without interdependence [37]. GHD prepubertal boys had a deficit in the cortical perimeter, not a deficit in length. These differences in geometry were seen only in the radius, possibly suggesting an increased tibial sensitivity to weight-bearing effects (i.e., mechanical loading due to locomotion) when compared with the radius [38], or an increased sensitivity of GHD bones to weight bearing compared to controls. Absence of differences may also be related to a lack of statistical power or some other unknown factor.

The differences in bone geometry that were seen in our cohort were not related to differences in lean or fat mass, as GHD subjects had similar body composition from DXA when compared to controls. No significant differences in vBMD, microarchitecture, bone strength, DXA measurements, body composition, or bone turnover markers were seen among the groups.

Animal models of GHD have shown deficits in bone geometry and size, and specifically in trabecular microarchitecture [36, 39–41]. Early treatment with GH in these mice was shown to fully restore trabecular microarchitecture [39] compared to other parameters that may only be restored partially. Our study did not find differences in trabecular microarchitecture, possibly because these differences might become more apparent later in puberty [42].

In contrast to our study, some prior studies using DXA in children have shown deficits in BMD in this population [2, 7–9]. However other studies have shown that when appropriate size corrections for body size were made, GHD was not associated with a significant decrease in BMD [43–47]. As DXA findings are influenced by bone size and DXA underestimates BMD when evaluating smaller bones, using HR-pQCT measurements allows for a more accurate assessment of true vBMD in these chil-

dren. Unfortunately, there are limited studies evaluating BMD using HR-pQCT in children this young, and therefore, a true reference range is not available.

No differences in body composition among the two groups were seen, though this is not surprising as alterations in body composition may only be seen in subjects with severe growth hormone deficiency [48]. We did however see associations between body composition, growth hormone, and IGF-1 levels. We found that in GHD subjects, lower peak GH levels were associated with increased fat mass, a finding supported by multiple studies showing that treatment with GH leads to a reduction in fat mass [7, 12, 49, 50]. We also found that IGF-1 was positively correlated with lean mass. Similar correlations between IGF-1 levels and lean body mass have been shown in pubertal girls [51] and children with cystic fibrosis [52]. Interestingly, IGF-1 levels in GHD subjects were also positively associated with cortical vBMD and cortical cross-sectional area, as well as whole-body areal BMD from DXA, similar to findings by Yang et al. [21] which showed that IGF-1 was positively correlated with total vBMD, cortical vBMD, and cortical area. These findings support the role of IGF-1 in muscle mass formation [53], bone health [54], and its role in the acquisition of peak bone mass [55].

The results of this study should be interpreted in the light of some limitations. It is a small, observational study and is meant to be used to generate further hypotheses in the field. It is cross-sectional, thus, associations do not prove causation. The diagnosis of GHD in childhood is challenging. We based the diagnosis of GHD on auxology, radiographic and biochemical data, and clinical judgment, which remain the foundation for the diagnosis. Our GHD subjects had a median GH level of 5.62 ng/mL, IGF-1 level of 140.93 ng/mL (123–275 ng/mL), delayed BA, and poor growth velocity. Thus, it is possible that had we limited our study to extreme cases of GHD, larger differences between the groups may have been seen. Interestingly, it has been shown that children with IGF-1 SDS < -2 did not differ significantly in anthropometric and body composition parameters from those with IGF-1 SDS ≥ -2, suggesting that IGF-1 in young prepubertal children may not necessarily be an adequate indication of growth hormone deficiency among children of short stature [48, 56].

Additionally, we studied only prepubertal boys in order to remove any effect of sex hormones, and therefore did not take into consideration the strong influence of maturation/puberty and other biological determinants of bone strength. Additionally, including girls in a future study may help evaluate the effect of other factors such as sex on pre-pubertal bone health. Our study was also not

large enough to stratify by ethnicity, which plays a role in BMD [24].

Our analysis using HR-pQCT scans was limited to peripheral sites and may not represent relationships between GHD and clinically relevant central sites. However, distal tibia parameters by HR-pQCT reflect the architecture of the central skeleton (i.e., proximal femur and lumbar spine) [57].

Despite the above limitations, our study was uniquely positioned to examine the influence of GHD on bone strength and parameters that underpin bone health in prepubertal GHD boys. HR-pQCT scans in young children can be technically difficult to perform due to the need for children to remain still and positioning problems secondary to small size. Nonetheless, we had minimal scan loss from motion artifact. The method used for supporting subjects with cushions and adding additional padding within the limb casts to mitigate limb movement, as well as focusing them with television shows worked well. Although children with GHD were found to have normal vBMD, HR-pQCT analysis revealed deficits in bone geometry that would have been missed by DXA. Short stature does not account for these deficits, as our findings persisted after controlling for height; limb length also does not account for our findings as length of radii were not significantly different between the two groups, and additionally, the same regions of interest in the radii were scanned across both groups. The findings appear to be driven primarily by a smaller cortical perimeter.

Clearly, growth hormone deficiency impacts bone size starting at a very young age. Larger studies are needed to evaluate the extent of these deficits, as well as the need for routine bone evaluation in this population. These findings lay the groundwork for investigations into the timing of earlier growth hormone replacement for prevention of these deficits and restoration of bone size, thereby enabling accrual of normal peak bone mass during adulthood.

### Statement of Ethics

All procedures involved in this research study were evaluated by Columbia University's Internal Review Board. Written informed consent was obtained from parents of all subjects.

Informed consent was obtained from a parent or legal guardian and assent was obtained for children greater than 7 years of age. The study was approved by the Institutional Review Board of Columbia University Medical Center.

### Disclosure Statement

The authors have no conflicts of interest to disclose.

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## Author Contributions

Tamar G. Baer was involved in all aspects of the study, including study concept, study design, recruitment of subjects, data analysis, and manuscript preparation. Sanchita Agarwal was involved in study design and specifically designed the scanning protocol for the HR-pQCT scans. She was also involved in data acquisition and interpretation, and manuscript preparation. Shaouxan Chen was

involved in statistical data analysis and interpretation and manuscript preparation. Codruta Chiuzan was involved in statistical data analysis and interpretation and manuscript preparation. Aviva Sopher was involved in study design, data acquisition, specifically interpretation of pediatric DXA scans, as well as manuscript preparation. Rachel Tao was involved in data acquisition and manuscript preparation. Abeer Hassoun was involved in study design, subject recruitment, and manuscript preparation. Elizabeth Shane was involved in study concept specifically with regard to HR-pQCT protocol design, and manuscript preparation. Ilene Fennoy was involved in study design, subject recruitment, data acquisition, specifically interpretation of pediatric DXA scans, as well as manuscript preparation. Sharon E. Oberfield was involved in study concept, study design, recruitment of subjects, and manuscript preparation. Patricia M. Vuguin is the principle investigator and was involved in all aspect of the study, including study concept, study design, recruitment of subjects, data analysis, and manuscript preparation.

## References

- Ohlsson C, Bengtsson BA, Isaksson OG, Andreassen TT, Sioström MC. Growth hormone and bone. *Endocr Rev*. 1998 Feb;19(1):55–79.
- Saggese G, Baroncelli GI, Bertelloni S, Barsanti S. The effect of long-term growth hormone (GH) treatment on bone mineral density in children with GH deficiency. Role of GH in the attainment of peak bone mass. *J Clin Endocrinol Metab*. 1996 Aug;81(8):3077–83.
- Davies JH, Evans BA, Gregory JW. Bone mass acquisition in healthy children. *Arch Dis Child*. 2005 Apr;90(4):373–8.
- Rizzoli R, Bianchi ML, Garabédian M, McKay HA, Moreno LA. Maximizing bone mineral mass gain during growth for the prevention of fractures in the adolescents and the elderly. *Bone*. 2010 Feb;46(2):294–305.
- Bonjour JP, Rizzoli R. Bone acquisition in adolescence. In: Marcus R, Feldman D, Kelsey J, editors. *Osteoporosis*. San Diego (CA): Academic Press; 1996. pp. 465–76.
- McCormack SE, Cousminer DL, Chesi A, Mitchell JA, Roy SM, Kalkwarf HJ, et al. Association Between Linear Growth and Bone Accrual in a Diverse Cohort of Children and Adolescents. *JAMA Pediatr*. 2017 Sep;171(9):e171769.
- Boot AM, Engels MA, Boerma GJ, Krenning EP, De Muinck Keizer-Schrama SM. Changes in bone mineral density, body composition, and lipid metabolism during growth hormone (GH) treatment in children with GH deficiency. *J Clin Endocrinol Metab*. 1997 Aug;82(8):2423–8.
- Baroncelli GI, Bertelloni S, Ceccarelli C, Saggese G. Measurement of volumetric bone mineral density accurately determines degree of lumbar undermineralization in children with growth hormone deficiency. *J Clin Endocrinol Metab*. 1998 Sep;83(9):3150–4.
- van der Sluis IM, Boot AM, Hop WC, De Rijke YB, Krenning EP, de Muinck Keizer-Schrama SM. Long-term effects of growth hormone therapy on bone mineral density, body composition, and serum lipid levels in growth hormone deficient children: a 6-year follow-up study. *Horm Res*. 2002;58(5):207–14.
- Rosén T, Wilhelmsen L, Landin-Wilhelmsen K, Lappas G, Bengtsson BA. Increased fracture frequency in adult patients with hypopituitarism and GH deficiency. *Eur J Endocrinol*. 1997;137(3):240–5.
- Vestergaard P, Jørgensen JO, Hagen C, Hoeck HC, Laurberg P, Rejnmark L, et al. Fracture risk is increased in patients with GH deficiency or untreated prolactinomas—a case-control study. *Clin Endocrinol (Oxf)*. 2002 Feb;56(2):159–67.
- Wüster C, Abs R, Bengtsson BA, Benmamer H, Feldt-Rasmussen U, Hernberg-Ståhl E, et al.; KIMS Study Group and the KIMS International Board. Pharmacia & Upjohn International Metabolic Database. The influence of growth hormone deficiency, growth hormone replacement therapy, and other aspects of hypopituitarism on fracture rate and bone mineral density. *J Bone Miner Res*. 2001 Feb;16(2):398–405.
- Bachrach LK, Gordon CM; SECTION ON ENDOCRINOLOGY. Bone Densitometry in Children and Adolescents. *Pediatrics*. 2016 Oct;138(4):e20162398.
- Cheung AM, Adachi JD, Hanley DA, Kendler DL, Davison KS, Josse R, et al. High-resolution peripheral quantitative computed tomography for the assessment of bone strength and structure: a review by the Canadian Bone Strength Working Group. *Curr Osteoporos Rep*. 2013 Jun;11(2):136–46.
- Nishiyama KK, Shane E. Clinical imaging of bone microarchitecture with HR-pQCT. *Curr Osteoporos Rep*. 2013 Jun;11(2):147–55.
- Sornay-Rendu E, Boutroy S, Munoz F, Delmas PD. Alterations of cortical and trabecular architecture are associated with fractures in postmenopausal women, partially independent of decreased BMD measured by DXA: the OFELY study. *J Bone Miner Res*. 2007 Mar;22(3):425–33.
- McCalden RW, McGeough JA, Barker MB, Court-Brown CM. Age-related changes in the tensile properties of cortical bone. The relative importance of changes in porosity, mineralization, and microstructure. *J Bone Joint Surg Am*. 1993 Aug;75(8):1193–205.
- Ulrich D, van Rietbergen B, Laib A, Ruegsegger P. The ability of three-dimensional structural indices to reflect mechanical aspects of trabecular bone. *Bone*. 1999 Jul;25(1):55–60.
- Chevalley T, Bonjour JP, van Rietbergen B, Ferrari S, Rizzoli R. Fractures during childhood and adolescence in healthy boys: relation with bone mass, microstructure, and strength. *J Clin Endocrinol Metab*. 2011 Oct;96(10):3134–42.
- Farr JN, Amin S, Melton LJ 3rd, Kirmani S, McCready LK, Atkinson EJ, et al. Bone strength and structural deficits in children and adolescents with a distal forearm fracture resulting from mild trauma. *J Bone Miner Res*. 2014 Mar;29(3):590–9.
- Yang H, Yan K, Yuping X, Zhang Q, Wang L, Gong F, et al. Bone microarchitecture and volumetric bone density impairment in young male adults with childhood-onset growth hormone deficiency. *Eur J Endocrinol*. 2019 Feb;180(2):145–53.
- Schweizer R, Martin DD, Schwarze CP, Binder G, Georgiadou A, Ihle J, et al. Cortical bone density is normal in prepubertal children with growth hormone (GH) deficiency, but initially decreases during GH replacement due to early bone remodeling. *J Clin Endocrinol Metab*. 2003 Nov;88(11):5266–72.

- 23 Gabel L, Macdonald HM, McKay HA. Sex Differences and Growth-Related Adaptations in Bone Microarchitecture, Geometry, Density, and Strength From Childhood to Early Adulthood: A Mixed Longitudinal HR-pQCT Study. *J Bone Miner Res*. 2017 Feb;32(2):250–63.
- 24 Gabel L, Macdonald HM, Nettlefold LA, McKay HA. Sex-, Ethnic-, and Age-Specific Centile Curves for pQCT- and HR-pQCT-Derived Measures of Bone Structure and Strength in Adolescents and Young Adults. *J Bone Miner Res*. 2018 Jun;33(6):987–1000.
- 25 Grimberg A, DiVall SA, Polychronakos C, Allen DB, Cohen LE, Quintos JB, et al.; Drug and Therapeutics Committee and Ethics Committee of the Pediatric Endocrine Society. Guidelines for Growth Hormone and Insulin-Like Growth Factor-I Treatment in Children and Adolescents: Growth Hormone Deficiency, Idiopathic Short Stature, and Primary Insulin-Like Growth Factor-I Deficiency. *Horm Res Paediatr*. 2016;86(6):361–97.
- 26 Zemel BS, Leonard MB, Kelly A, Lappe JM, Gilsanz V, Oberfield S, et al. Height adjustment in assessing dual energy x-ray absorptiometry measurements of bone mass and density in children. *J Clin Endocrinol Metab*. 2010 Mar;95(3):1265–73.
- 27 Agarwal S, Rosete F, Zhang C, McMahon DJ, Guo XE, Shane E, et al. In vivo assessment of bone structure and estimated bone strength by first- and second-generation HR-pQCT. *Osteoporos Int*. 2016 Oct;27(10):2955–66.
- 28 Manske SL, Zhu Y, Sandino C, Boyd SK. Human trabecular bone microarchitecture can be assessed independently of density with second generation HR-pQCT. *Bone*. 2015 Oct;79:213–21.
- 29 Sode M, Burghardt AJ, Pialat JB, Link TM, Majumdar S. Quantitative characterization of subject motion in HR-pQCT images of the distal radius and tibia. *Bone*. 2011 Jun;48(6):1291–7.
- 30 Boutroy S, Van Rietbergen B, Sornay-Rendu E, Munoz F, Bouxsein ML, Delmas PD. Finite element analysis based on in vivo HR-pQCT images of the distal radius is associated with wrist fracture in postmenopausal women. *J Bone Miner Res*. 2008 Mar;23(3):392–9.
- 31 Müller R, Rüeegsegger P. Three-dimensional finite element modelling of non-invasively assessed trabecular bone structures. *Med Eng Phys*. 1995 Mar;17(2):126–33.
- 32 Macneil JA, Boyd SK. Bone strength at the distal radius can be estimated from high-resolution peripheral quantitative computed tomography and the finite element method. *Bone*. 2008 Jun;42(6):1203–13.
- 33 Pistoia W, van Rietbergen B, Lochmüller EM, Lill CA, Eckstein F, Rüeegsegger P. Estimation of distal radius failure load with micro-finite element analysis models based on three-dimensional peripheral quantitative computed tomography images. *Bone*. 2002 Jun;30(6):842–8.
- 34 Duan Y, Parfitt A, Seeman E. Vertebral bone mass, size, and volumetric density in women with spinal fractures. *J Bone Miner Res*. 1999 Oct;14(10):1796–802.
- 35 Seeman E. Periosteal bone formation—a neglected determinant of bone strength. *N Engl J Med*. 2003 Jul;349(4):320–3.
- 36 Kristensen E, Hallgrímsson B, Morck DW, Boyd SK. Timing of growth hormone treatment affects trabecular bone microarchitecture and mineralization in growth hormone deficient mice. *Bone*. 2010 Aug;47(2):295–300.
- 37 Bonaretti S, Majumdar S, Lang TF, Khosla S, Burghardt AJ. The comparability of HR-pQCT bone measurements is improved by scanning anatomically standardized regions. *Osteoporos Int*. 2017 Jul;28(7):2115–28.
- 38 Mikkola TM, Sipilä S, Rantanen T, Sievänen H, Suominen H, Kaprio J, et al. Genetic and environmental influence on structural strength of weight-bearing and non-weight-bearing bone: a twin study. *J Bone Miner Res*. 2008 Apr;23(4):492–8.
- 39 Chaudhry AA, Castro-Magana M, Aloia JF, Yeh JK. Differential effects of growth hormone and alpha calcidol on trabecular and cortical bones in hypophysectomized rats. *Pediatr Res*. 2009 Apr;65(4):403–8.
- 40 Kristensen E, Hallgrímsson B, Morck DW, Boyd SK. Microarchitecture, but not bone mechanical properties, is rescued with growth hormone treatment in a mouse model of growth hormone deficiency. *Int J Endocrinol*. 2012;2012:294965.
- 41 Chen MM, Yeh JK, Aloia JF. Histologic evidence: growth hormone completely prevents reduction in cortical bone gain and partially prevents cancellous osteopenia in the tibia of hypophysectomized rats. *Anat Rec*. 1997 Oct;249(2):163–72.
- 42 Gilsanz V, Gibbens DT, Roe TF, Carlson M, Senac MO, Boechat MI, et al. Vertebral bone density in children: effect of puberty. *Radiology*. 1988 Mar;166(3):847–50.
- 43 Höglér W, Shaw N. Childhood growth hormone deficiency, bone density, structures and fractures: scrutinizing the evidence. *Clin Endocrinol (Oxf)*. 2010 Mar;72(3):281–9.
- 44 Höglér W, Briody J, Moore B, Lu PW, Cowell CT. Effect of growth hormone therapy and puberty on bone and body composition in children with idiopathic short stature and growth hormone deficiency. *Bone*. 2005 Nov;37(5):642–50.
- 45 Maheshwari HG, Bouillon R, Nijs J, Oganov VS, Bakulin AV, Baumann G. The Impact of congenital, severe, untreated growth hormone (GH) deficiency on bone size and density in young adults: insights from genetic GH-releasing hormone receptor deficiency. *J Clin Endocrinol Metab*. 2003 Jun;88(6):2614–8.
- 46 Benbassat CA, Eshed V, Kamjin M, Laron Z. Are adult patients with Laron syndrome osteopenic? A comparison between dual-energy X-ray absorptiometry and volumetric bone densities. *J Clin Endocrinol Metab*. 2003 Oct;88(10):4586–9.
- 47 Woods KA, Camacho-Hübner C, Bergman RN, Barter D, Clark AJ, Savage MO. Effects of insulin-like growth factor I (IGF-I) therapy on body composition and insulin resistance in IGF-I gene deletion. *J Clin Endocrinol Metab*. 2000 Apr;85(4):1407–11.
- 48 Matusik P, Klesiewicz M, Klos K, Stasiulewicz M, Barylak A, Nazarkiewicz P, et al. Baseline Body Composition in Prepubertal Short Stature Children with Severe and Moderate Growth Hormone Deficiency. *Int J Endocrinol*. 2016;2016:4563721.
- 49 Aurenzans Clemente E, Samper Villagrasa P, Ayerza Casas A, Ruiz Frontera P, Bueno Lozano O, Moreno Aznar LA, et al. [Effects of growth hormone treatment on anthropometrics, metabolic risk, and body composition variables in small for gestational age patients]. *An Pediatr (Barc)*. 2017 May;86(5):240–8.
- 50 Kuromaru R, Kohno H, Hara T. Changes in adiposity and excess body weight correlate with growth responses but not with decreases in low-density lipoprotein cholesterol levels during GH treatment in GH-deficient children. *Clin Endocrinol (Oxf)*. 2002 Jun;56(6):799–803.
- 51 Cirmanova V, Zofkova I, Kasalicky P, Lanska V, Bayer M, Starka L, et al. Hormonal and bone parameters in pubertal girls. *Physiol Res*. 2017 Sep;66(3 Supplementum 3):S419–24.
- 52 Sermet-Gaudelus I, Souberbielle JC, Azhar I, Ruiz JC, Magnine P, Colomb V, et al. Insulin-like growth factor I correlates with lean body mass in cystic fibrosis patients. *Arch Dis Child*. 2003 Nov;88(11):956–61.
- 53 Clemmons DR, Underwood LE. Role of insulin-like growth factors and growth hormone in reversing catabolic states. *Horm Res*. 1992;38 Suppl 2:37–40.
- 54 Yakar S, Rosen CJ, Beamer WG, Ackert-Bicknell CL, Wu Y, Liu JL, et al. Circulating levels of IGF-1 directly regulate bone growth and density. *J Clin Invest*. 2002 Sep;110(6):771–81.
- 55 Langlois JA, Rosen CJ, Visser M, Hannan MT, Harris T, Wilson PW, et al. Association between insulin-like growth factor I and bone mineral density in older women and men: the Framingham Heart Study. *J Clin Endocrinol Metab*. 1998 Dec;83(12):4257–62.
- 56 Galluzzi F, Quaranta MR, Salti R, Saieva C, Nanni L, Seminara S. Are IGF-I and IGF-BP3 useful for diagnosing growth hormone deficiency in children of short stature? *J Pediatr Endocrinol Metab*. 2010 Dec;23(12):1273–9.
- 57 Liu XS, Cohen A, Shane E, Yin PT, Stein EM, Rogers H, et al. Bone density, geometry, microstructure, and stiffness: relationships between peripheral and central skeletal sites assessed by DXA, HR-pQCT, and cQCT in premenopausal women. *J Bone Miner Res*. 2010 Oct;25(10):2229–38.